ANTIOXIDANTS SELENOMETHIONINE AND D-PANTETHINE DIFFERENTIALLY AFFECT DOXORUBICIN’S ACTION ON GLUTATHIONE SYSTEM IN HUMAN LEUKEMIA CELLS VARYING IN THEIR RESISTANCE TO CHEMOTHERAPY IN VITRO

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Rapid development of multiple drug resistance and occurrence of negative side effects in cancer patients arising at the treatment belong to the main problems in cancer chemotherapy. Recently, it was shown that specific antioxidants (selenomethionine – SeMet and D-pantetheine – D-Pt) possessed nephro-, myelo- and hepatoprotective activity at doxorubicin’s (Dx) action in tumor-bearing mice. Besides, these antioxidants inhibited a cytotoxic action of Dx toward chemotherapy-sensitive tumor cells, and enhanced the cytotoxic effect of this drug toward selected drug-resistant tumor cell lines (e.g. HL-60/vinc, HL-60/adr), while in other such lines (e.g. HCT-116/Bax(−/−), HCT-116/p53), it was not effective.

The aim of present study was to investigate the molecular mechanisms of the revealed difference in the action of SeMet and D-Pt toward cytotoxic effects of Dx in tumor cells varying in drug resistance. Human leukemia cells of HL-60/wt line and its drug-resistant sublines HL-60/adr (overexpression of MRP-1) and HL-60/vinc (overexpression of P-gp) were used in this study.

Treatment of cells with Dx led to the versatile action of this drug on the level of glutathione in each of the studied cell line and sublines. HL-60/wt cells were characterized...
by 8-fold lower GSH level under Dx treatment compared to control, while in HL-60/vinc and HL-60/adr cells GSH level was increased 2.2- and 8.2-fold (compared to untreated cells), correspondingly. The use of doxorubicin also led to significant rearrangement of GSSG/GSH ratio in these cell lines, leading to 2-fold elevation of GSSG level HL-60/vinc cells, and 2.5-fold decrease of this index in HL-60/adr cells.

We have shown that a combined effect of SeMet or D-Pt on the background of the cytotoxic action of doxorubicin on HL-60/vinc cells is accompanied by a 2-fold decrease in both oxidized and reduced glutathione levels. Such an effect of these antioxidants can serve as an explanation of their sensitizing effect on the cells of the HL-60/vinc subline under Dx's action which we observed earlier. It should be noted that treatment with Dx led to a 2.5-fold increase in the activity of glutathione-S-transferase in the leukemia cells of HL-60/vinc subline. The antioxidants effectively reduced this indicator. SeMet and D-Pt differentially affected the activity of glutathione-S-transferase in HL-60/adr cells.

In conclusion, our data demonstrate an important role of the antioxidants on the functional state of the glutathione system in tumor cells that differ in their drug resistance. The obtained results suggest an important role of glutathione-S-transferase in modulation of cancer drug resistance that is caused by P-glycoprotein overexpression, but not by the overexpression of MRP-1 protein. Selenomethionine and D-pantethine effectively inhibit this enzyme, thus, sensitizing P-gp overexpressing cells towards the action of doxorubicin. This event is accompanied by further decrease in GSH and GSSG levels in these cells, thus sensitizing them to Dx action. Further studies of the molecular mechanisms underlying this phenomenon are in progress.

**Keywords:** doxorubicin, antioxidants, glutathione, glutathione peroxidase, glutathione reductase, glutathione-S-transferase

**INTRODUCTION**

Chemotherapy is the main treatment method of cancer today. But traditional anticancer drugs don’t always provide a complete remission. Besides a lot of antitumor drugs have some side effects, because they affect both malignant and normal cells of organism. In addition, one of the main problem of modern chemotherapy is rapid development of the multidrug resistance (MDR) of malignant cells. It significantly reduces the effectiveness of treatment of the oncological patients. This phenomenon is associated with an elevated expression of membrane-bound ATP binding cassette transporter proteins (ABC transporters) that pump out the anticancer drugs from the malignant cells [6]. The ABC-transporters play an important role in human organism, protecting healthy tissues (in particular, liver, small and large intestines and kidney) from the action of various xenobiotics [1]. However, their overexpression in the malignant cells significantly reduces the sensitivity of tumors to chemotherapy. P-gp (P-glycoprotein), MRP-1 (multidrug-resistance associated protein) and BCRP (breast cancer resistance protein) are the most important members of this family, and, thus, development of novel efficient inhibitors of these proteins remains an actual task in modern oncology [4, 5, 13].

The solution of the main problems of chemotherapy is possible using specific agents which reduce the negative side effects of antitumor drugs and by which the phenomenon of multiple drug resistance could be overcome. In previous studies, we have shown that antioxidants selenomethionine (SeMet) and D-pantethine (D-Pt) are promising candidates for the role of such agents. It was found that antioxidant supplements possess
nephro-, myelo- and hepatoprotective activity toward doxorubicin (Dx) action on the organism of tumor-bearing mice. Also, these antioxidants partially sensitized drug-resistant tumor cells toward cytotoxic action of the doxorubicin (Dx) [15–17]. However, the molecular mechanisms underlying such effects of selenomethionine and D-pantethine remain unexplored.

One of the mechanism for the emergence of multiple drug resistance is the glutathione system. It is known that the antioxidants are potential modulators of cellular glutathione system that is often involved in regulation of drug resistance in tumor cells [23, 24]. Glutathione is a biologically active tripeptide. Sulphydryl group of this thiol (R–SH) interacts with the reactive group of the chemotherapeutic drug to form a conjugate. Such conjugates are less active and are removed from the cell by ATP binding cassette transporter proteins. Chemical interactions between glutathione and antitumor compounds are catalyzed by glutathione-S-transferase enzymes. Glutathione-S-transferases (GSTs) are a family of detoxification enzymes that catalyse the conjugation of glutathione (GSH) to a wide variety of endogenous and exogenous compounds.

Literature data confirm an important role of reduced glutathione (GSH) and glutathione-S-transferase in transport and accumulation of anticancer drugs in drug-resistant tumor cells. Thus, it was demonstrated that that elevated levels of GSH, together with increased activities of glutathione-S-transferase (GST) or peroxidase, may protect cells from cytotoxic drugs (platinum compounds and anthracyclines). Conversely, depletion of the cellular glutathione (GSH) levels inhibits the transport of the drug through inhibition of the transport of glutathione S-conjugates [22, 24]. It is known that cytosolic GSTs of some classes play a regulatory role in the mitogen-activated protein (MAP) kinase pathway that participates in cellular survival and death signals. So GSTs perform their function by direct detoxification or act as inhibitors of the MAP kinase pathway. Such an effect of GSTs on the MAP kinase pathway explains why in many cases the drugs used to select for resistance don’t conjugate with GSH and are not substrates for GSTs [22, 23].

So, the aim of this study was to address potential mechanisms of the effect of antioxidant compounds on the functional state of the glutathione system in tumor cells that differ in their drug resistance.

**MATERIALS AND METHODS**

Human leukemia HL-60 cell line and its drug-resistant sublines HL-60/adr (overexpression of MRP-1) and HL-60/vinc (overexpression of P-glycoprotein) were kindly provided by prof. Walter Berger (cell culture collection at the Institute of Cancer Research, Vienna Medical University, Austria).

Cells were cultured in the RPMI-1640 medium (Sigma-Aldrich, St. Louis, USA) supplemented with 10% fetal calf serum (Sigma-Aldrich, St. Louis, USA), 50 µg/ml streptomycin (Sigma-Aldrich, St. Louis, USA), and 50 units/ml penicillin (Sigma-Aldrich, St. Louis, USA) in 5% CO₂ humidified atmosphere at 37 °C. During experiments, cells (1×10⁶ per ml) were incubated for 24 h in 6-well tissue culture plates (Greiner Bio-one, Frickenhausen, Germany), and the substances was added at various concentrations (doxorubicin in a dose of 1 µM for HL-60/wt cells, 5 µM for HL-60/vinc cells and 10 µM for HL-60/adr cells). The concentration of the antioxidants was the same for all studied cell lines, namely 0.2 µM for selenomethionine and 10 µM for D-pantethine. Cytotoxic effects of Dx and antioxidants were evaluated under Evolution 300 Trino microscope
(Delta Optical, Nowe Osiny, Poland) by staining the dead cells with trypan blue dye (0.1%). After that, cells were centrifuged at 1,500 rpm for 5 min, and cell suspension was washed twice with a phosphate buffered saline.

The measurement of glutathione was carried out using the Ellman’s Reagent (DTNB – 5,5′-Dithiobis-2-Nitrobenzoic Acid), which forms a colored complex with a maximal absorption at 412 nm with SH-groups at pH 8.0 and with SS-groups at pH 10.5. Cell suspension was deproteinized with 0.4 ml of 50% sulfosalicylic acid, centrifuged, and the resulting supernatant was neutralized to pH 7.6 by the addition of 0.1 ml of cooled 2.5 M K₃PO₄. After additional centrifugation, the entire supernatant was mixed with 2 ml of phosphate buffer (pH 8.0) containing 0.004% Ellman’s reagent. The optical density of the solution was measured by a spectrophotometer SF-46 (LOMO, Russian Federation) at a wavelength of 412 nm. After that, pH in the cuvette was quickly increased to 10.5, and a marked increase in absorption was observed. The content of the oxidized glutathione (GSSG) was calculated as the difference between absorption of complex of Ellman’s reagent at pH 10.5 and pH 8.0 [19].

The glutathione reductase (GR, EC 1.8.1.7) activity was determined as the oxidation rate of NADPH by the oxidized glutathione (GSSG)n that was recorded spectrophotometrically. 100 µl of supernatant was added to 2.4 ml of 0.01 M phosphate buffer (pH 7.4) containing 1 mM EDTA, 2 mM GSSG and 0.2 mM NADPH, and the absorption decrease was recorded at 340 nm [3]. The glutathione transferase (GST, EC 2.5.1.18) activity was determined as the rate of formation of a conjugate between reduced glutathione (GSH) and CDNB (1-Chloro-2,4-dinitrobenzene). 50 µl of samples were added to 2 ml of 0.1 M phosphate buffer (pH 6.5) containing 2.5 mM of GSH and 1.0 mM of CDNB. Then the absorption increase at 340 nm was recorded using a molar absorption coefficient (9.6 mm−1 cm−1) [18]. The glutathione peroxidase (GPx, EC 1.11.1.9) activity was measured by the colorimetric method using a decrease of GSH concentration under reaction with several types of hydroperoxides. For determination of the GPx activity of the method of V. M. Moin [11]. A solution of tert-butyl hydroperoxide (20 mM) was added to the reaction mixture containing 100 µl cell suspension (1:29), 830 µl Tris buffer (0.1 M, pH 8.5 with addition of 6 mmol EDTA, 9.96 µmol sodium azide and 4.8 mmol GSH). The amount of GSH consumed in the process of interaction between the enzyme and the substrate was determined spectrophotometrically at 412 nm upon the appearance of a colored thionitrofenyl product of 5,5′-dithiobis-2-nitrobenzoic acid, taking into account the molar extinction coefficient (13 100) of the Ellman’s reagent.

All experiments were performed in triplicate and repeated 3 times. Statistical analysis of data was conducted in GraphPad Prism Software (GraphPad Software, Inc) using Student’s t-test. Statistical significance was set at P≤0.05.

RESULTS AND DISCUSSION

In order to reveal potential interrelations between glutathione level and sensitivity of tumor cells to anticancer drugs, cellular content of oxidized (GSSG) and reduced glutathione (GSH) was studied in human leukemia HL-60/wt cell line and its drug-resistant sublines. It was found that basal GSH level was identical both in HL-60/wt cells and its vincristine-resistant HL-60/vinc subline (overexpressing P-glycoprotein), while in doxorubicin-resistant cells of HL-60/adr subline (overexpression of MRP-1) this index was 2-fold lower (Fig.1, Table 1). On contrary, basal level of oxidized glutathione (GSSG) was increased 2-fold in HL-60/adr subline, while HL-60/vinc cells were characterized by
6-fold lower level of GSSG compared to the parental cells. So, the cells of each of the studied cell lines are characterized by different basal levels of both oxidized and reduced glutathione, leading to different GSH/GSSG ratios – 0.34 in HL-60/wt, 2.2 in HL-60/vinc and 0.06 in HL-60/adr (Fig. 1, Table 1). Such a difference in these indices may be explained by overexpression of ABC-transporter proteins in drug-resistant tumor cells, which can utilize glutathione for their own needs. In particular, it is known that overexpression of MRP-1 protein is closely related to an increased level of glutathione in tumor cells [24]. GSH forms adducts with anticancer drugs which are then pumped out from cells by the MRP-1 protein. At the same time, the expression of P-glycoprotein is not directly related to the level of cellular glutathione [24] that might explain another pattern of its level in the cells of the HL-60/vinc subline.

The next aim of this work was to study if specific modulators of cellular glutathione system (selenomethionine, SeMet and D-pantethine, D-Pt) might sensitize drug-resistant tumor cells to the action of various anticancer drugs. It is known that selenium is a coenzyme of glutathione peroxidase and plays an important role in the regulation of the glutathione system [20]. D-pantethine is the precursor of coenzyme A, which is involved in most metabolic processes in the cell [2, 12]. Both compounds led to major reduction of GSH levels in HL-60/wt cells, insignificantly influenced on this index in HL-60/vinc cells and massively (3- and 6-fold, correspondingly) enhanced it in HL-60/adr cells. On contrary, there was no impact of SeMet and D-Pt on GSSG levels in HL-60/wt and HL-60/vinc cells, but in HL-60/adr cells it was significantly (2- and 2.4-fold, correspondingly) decreased (Fig. 1, Table 1). These data clearly indicate on activation of
MRP-1 protein in HL-60/adr cells under their treatment with SeMet and D-Pt, that is accompanied by significant rearrangement in cellular GSH/GSSG ratio.

In order to confirm this suggestion, we studied changes in cellular GSH and GSSG levels under impact of well-known anticancer drug doxorubicin (Dx) that is a typical classic substrate of ABC-transporter proteins [21]. Treatment of cells with Dx led to the versatile action of this drug on the level of glutathione in each of the studied cell line and sublines. HL-60/wt cells were characterized by 8-fold lower GSH level under Dx treatment compared to control, while in HL-60/vinc and HL-60/adr cells GSH level was increased 2.2- and 8.2-fold (compared to the untreated cells), correspondingly (Fig. 1, Table 1). These results clearly indicate an activation of ABC-transporter proteins in MDR cell lines under Dx treatment that requires reduced glutathione for their activity. This event also led to a significant rearrangement of GSSG/GSH ratio in these cell lines, leading to 2-fold elevation of GSSG.

Table 1. Impact of selenomethionine (SeMet) and D-pantethine (D-Pt) on the level of glutathione (reduced – GSH and oxidized – GSSG) and on the activity of glutathione-S-transferase (GST) in human leukemia cells of the HL-60/wt line and its resistant subline HL-60/adr and HL-60/vinc under doxorubicin (Dx) action

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<tr>
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<td>GSSG</td>
<td>GSH</td>
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<td>↓7x</td>
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<td>Dx</td>
<td>↑1.5x</td>
<td>↓7.3x</td>
<td>↑1.4x</td>
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<tr>
<td>Dx + D-Pt</td>
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<td>↑1.4x (18x)#</td>
<td>↑1.1x (no effect)#</td>
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<tr>
<td>Dx + SeMet</td>
<td>no effect (1.5x)#</td>
<td>↓5.5x (no effect)#</td>
<td>↑1.3x (no effect)#</td>
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Comments: # – compared to HL-60/vinc, control; ▼ – compared to HL-60/vinc, Dx;▲ – compared to HL-60/adr, control; • – compared to HL-60/adr, Dx; ■ – compared to HL-60/wt, Dx

Примітки: # – порівняно з клітинами сублінії HL-60/vinc, контроль; ▼ – порівняно з клітинами сублінії HL-60/vinc, Dx;▲ – порівняно з клітинами сублінії HL-60/adr, контроль; • – порівняно з клітинами сублінії HL-60/adr, Dx; ■ – порівняно з клітинами лінії HL-60/wt, Dx
level HL-60/vinc cells, and 2.5-fold decrease of this index in HL-60/adr cells. Concluding, enhancement of cellular GSH level and loss of GSSG level under Dx treatment indicates on increased tumor cell resistance to this drug, while a decrease in GSH level, accompanied with an increase of GSSG level demonstrates the opposite situation.

We have shown that a combined effect of SeMet or D-Pt on the background of the cytotoxic action of doxorubicin on HL-60/vinc cells is accompanied by a 2-fold decrease in both oxidized and reduced glutathione levels (Fig. 1, Table 1). Such an effect of these antioxidants can serve as an explanation of their sensitizing effect on the cells of the HL-60/vinc subline under Dx’s action, which we observed earlier [15].

For revealing the molecular mechanisms underlying this phenomenon, the impact of doxorubicin, SeMet and D-Pt on the activity of key enzymes (glutathione peroxidase (GPx), glutathione reductase (GR) and glutathione-S-transferase (GST) of the glutathione system was studied. We did not detect significant changes in the activity of glutathione peroxidase in the studied cell line and sublines. There was no effect of the antioxidants and Dx on these indicators as well (Fig. 2).

![Fig. 2. Impact of joint application of selenomethionine (SeMet) or D-pantethine (D-Pt) with doxorubicin (Dx) on the activity of glutathione peroxidase (GPx) and glutathione reductase (GR) in human leukemia cells of the HL-60/wt line and its resistant subline HL-60/adr and HL-60/vinc (24 h incubation). ** – P <0.01; **** – P <0.0001 (vs control); • – P <0.05; •• – P <0.01 (vs doxorubicin)](image)

The glutathione reductase activity was considerably (10 times) reduced under the action of Dx on wild-type leukemia cells (HL-60/wt), and it was partially recovered under the action of this drug in a combination with SeMet or D-Pt (Fig. 2). Thus, we found a positive correlation between the activity of this enzyme and the level of reduced glutathione under the influence of Dx that is consistent with the literature data [7]. At the same time, the cells of HL-60/vinc and HL-60/adr sublines showed a completely different activity of this enzyme (Fig. 2). In particular, the overexpression of the MRP-1 protein in HL-60/adr cells was accompanied by a 3-fold increase in the activity of glutathione...
reductase. This activity was lowered twice under the action Dx, while both antioxidants did not demonstrate a modulating effect here. Basal activity of glutathione reductase in HL-60/vinc cells did not differ from this level in the wild-type cells (HL-60/wt), while Dx treatment led to a 2-fold increase in the activity of this enzyme. SeMet and D-Pt partially normalized the level of that indicator (Fig. 2).

Most significant changes were detected at analysis of the activity of glutathione-S-transferase in tumor cells (Fig. 3, Table 1). In both drug-resistant sublines of the leukemia cells, there was a 2-fold increase of the activity of this enzyme. These results are consistent with the literature data about the glutathionylation of xenobiotics and drugs as an important step before their excretion from cells by the ABC-transporters [8, 10, 23]. Treatment of HL-60/vinc cells with Dx led to a 2.5-fold increase of glutathione-S-transferase activity, which also positively correlated with 2-fold increase of GSH level in these cells. Both SeMet or D-Pt lowered GST activity to a basal level, thus, leading to subsequent decrease in GSH and GSSG levels (Fig. 3, Table 1). These data indicate a pronounced inhibitory effect of both antioxidants on the activity of glutathione-S-transferase that might be explained by their ability to enhance the action of the doxorubicin on the malignant cells resistant to chemotherapy (showed by us in [15]).

It should be stressed that similar effect of SeMet and D-Pt on the activity of glutathione-S-transferase in drug-resistant HL-60/adr cells was absent, probably due to another mechanism of drug resistance in these cells. Moreover, a co-treatment of HL-60/adr cells with SeMet and Dx led to an enhancement of GST activity, accompanied by increase of GSH levels in these cells (Fig. 1, Fig. 3). Thus, SeMet and D-Pt can sensitize to chemotherapy only the P-gp-overexpressing tumor cells, and had a weak influence on MRP-1 overexpressing cancer cell lines.

**Fig. 3.** Impact of selenomethionine and D-pantethine on the activity of glutathione-S-transferase (GST) in human leukemia cells of the HL-60/wt line and its resistant subline HL-60/adr (MRP-1 +) and HL-60/vinc (P-gp +) under the action of doxorubicin (24 h incubation).** – P <0.01 (vs control); *** P <0.001; **** – P <0.0001 (vs doxorubicin).

Рис. 3. Вплив селенометіоніну (SeMet) і D-пантетину (D-Pt) на активність глутатіон-S-трансферази (GST) у клітинах лейкозу людини лінії HL-60/wt і її резистентних субліній HL-60/adr (MRP-1 +) та HL-60/vinc (P-gp +), які піддавалися обробці доксорубіцином (Dx) (24 год інкубації). ** – P <0.01 (порівняно з контролем); *** – P <0.001; **** – P <0.0001 (порівняно з доксорубіцином)
CONCLUSIONS

The obtained results suggest an important role of glutathione-S-transferase in modulation of cancer drug resistance that is caused by P-glycoprotein overexpression, but not by the overexpression of MRP-1 protein. Selenomethionine and D-pantethine effectively inhibit this enzyme, thus, sensitizing P-gp overexpressing cells towards the action of doxorubicin. This event is accompanied by further decrease in GSH and GSSG levels in these cells, thus sensitizing them to Dx action. Further studies of the molecular mechanisms underlying this phenomenon are in progress.

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Завдання дослідження було вивчення потенційних молекулярних механізмів впливу антиоксидантних сполук на функціональний стан системи глутатіону в пухлинних клітинних лініях (HL-60/wt, HL-60/adr, HL-60/vinc) та їх резистентних субліній (HCT-116/Bax(−/−), HCT-116/p53). Використання доксорубіцину призвело до суттєвої перебудови співвідношення GSSG / GSH у клітинах цих субліній, що призвело до 2-кратного підвищення

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рівня GSSG у клітинах лінії HL-60/vinc та 2,5-кратного зменшення цього показника в клітинах лінії HL-60/adr. Варто зазначити, що вплив Dx на клітини сублінії HL-60/vinc призвів до 2,5-кратного збільшення активності глутатіон-S-трансферази. Важливим є те, що антиоксиданти ефективно знизили цей показник, проте вони не продемонстрували подібного впливу на активність глутатіон-S-трансферази в клітинах сублінії HL-60/adr.

Отже, наші дані демонструють важливу роль антиоксидантів у регуляції функціонального стану системи глутатіону в пухлинних клітинах, що характеризуються різними механізмами стійкості до ліків.

**Ключові слова:** доксорубіцин, антиоксиданти, глутатіон, глутатіонпероксидаза, глутатіонредуктаза, глутатіон-S-трансфераза